

Effect of *Ulva lactuca* on Indole-3-acetic acid, Siderophores and Phosphatase Production by *Azospirillum brasilense* NH Growing in Seawater as Natural Medium

El Hafid Nabti*

Laboratory of Mastery of Renewable Energies, University of Bejaia, Targa Ouzemmour, 06000 Bejaia, Algeria.

Received: 28 Jun. 2012, Revised: 12 Nov. 2012, Accepted: 20 Nov. 2012

Published online: 1 May. 2013

Abstract: In this paper we aim to synthesize a new culture medium similar to the seawater, and test the osmotolerant strain *Azospirillum brasilense* NH for IAA and siderophores-production and phosphate solubilization in presence of a natural osmoprotectant *Ulva lactuca* (marine alga). This is in order to reveal the capacity of this strain to restore a plant growth irrigated with seawater or brackish water in the future. *Azospirillum brasilense* NH a new isolate from Algerian soil known as an osmotolerant bacterium was tested with natural medium containing seawater (32g/l NaCl) in presence of ethanol algal extracts and absence of other carbon source such as malic acid, for growth and indole acetic acid production. The results obtained showed that bacterium NH increased its growth at 25% of algal extracts and produced high concentration of auxin phytohormone indole acetic acid from tryptophan (64.61 µg/ml) in absence of NH₄Cl. Siderophores production and inorganic phosphates solubilization were tested then positive results were obtained. These results demonstrated that *A. brasilense* NH get its natural environment in seawater in presence of algal extracts. So, the capability of the strain to keep the Plant Growth Promoting Rhizobacteria characters under seawater using *U. lactuca* to save its growth, let us think about an eventual application as a biofertilizer in agricultural soils irrigated with saline water or menaced with salinity. As we demonstrated already, that this strain stimulated a wheat growth under saline conditions, but, here, we have to look for an application of the bacterium in large scale. Because, now we need to save the crop invaded by seawater and other saline waters.

Keywords: *Azospirillum brasilense* NH, Seawater -*Ulva lactuca*, IAA-production, Siderophores, phosphatase.

1 Introduction

In our time, aridity and water missing become factors threatening soil. Many countries suffer from the consequences of these biotic stress. Algeria is one of the countries where the aridity touches more than three quarters of its surface. So, the use of osmotolerant bio-fertilizer bacteria (e.g., *Azospirilla*) and marine algae (e.g., *Ulva*), could provide a solution for problem.

The selection of plant growth promoting rhizobacteria (PGPR) will be a good idea in the application of the biological fertilizer. The effect of *Azospirillum* on plant growth has been intensively studied, as reported by many authors [3,6,11,12]. *Azospirillum* Sp. is known by its improvement of crop yield with inoculation of plants roots, this plant responses to *Azospirillum* inoculation is

attributed to the indole-3-acetic acid (IAA) production by these bacteria [2,16].

Various authors suggested direct promoting mechanisms in addition to IAA production [17], and osmotic stress response in plants [1,4,12], phosphate solubilization [20] and siderophore production [18].

Aridity and lack water in the soil inhibit the plant growth. Therefore, our study focused on synthesis of medium containing Seawater which could be a sustainable solution to fight against water missing, in one hand, and the combination of the marine alga *Ulva lactuca* with the halophilic and osmotolerant PGPR to challenge the osmotic stress due to the salinity pressure of Seawater, in other hand. So, the osmoprotective effect of the alga

* Corresponding author e-mail: elhnabti1977@yahoo.fr

allows the restoration of IAA (auxin-phytohormone) synthesis; siderophores production and inorganic phosphate solubilization.

The algal extracts contain high amounts of different betaines, amino-acids, proteins, dimethylsulphoniopropionate and nitrogen source [7]. IAA is the most important naturally occurring of plant growth and development [17]. In this paper we tried to show the effect of marine *U. lactuca* on restoration of production of IAA in Seawater.

2 Materials and Methods

Microorganism: *Azospirillum brasilense* NH isolated from Algeria [13]. Vegetal material: the marine alga *Ulva lactuca* isolated from the Gulf of Bejaia in Algeria. Ethanol algal extracts was obtained according to [7]. Semisolid medium contains: 1l of Seawater preliminary autoclaved (110°C for 30 min), and 5ml 0.5 % alcoholic solution of bromothymol blue (BBT), algal extracts (25 %) and agar (1.75 g). Another medium was prepared without algal extracts as control. pH was adjusted with KOH at 6.8. Media were inoculated with 1ml of bacterial solution of *A. brasilense* NH then, incubation conditions were monitored at 32°C for 72 h. Solid medium was prepared with same composition cited above, but without BBT. Agar was added (15g/l). One colony was removed from the semisolid medium containing algal extracts to inoculate the solid medium. A control was used without algal extracts.

2.1 Indole Acetic Acid synthesis

The protocol was inspired from [5], but the medium is modified here by using Seawater instead of distilled water. Bacteria of *A. brasilense* NH were grown in the liquid medium prepared with water Sea, algal extracts and tryptophan (0.5 mg/l), but without NH₄Cl under shaking conditions (100 rpm) at 32°C to the late stationary phase. The same medium was prepared but without algal extracts as control and inoculated with *A. brasilense* NH. Culture supernatant was obtained by centrifugation at 10.000 rpm for 15 min. Determination of concentration of IAA produced was performed according to [5] and [13].

2.2 Phosphate solubilization

The NBRIP medium was utilized [14] containing the following composition in g/l of Seawater: Glucose (10 g), Ca₃(PO₄)₂ (5g), MgCl₂ x 6H₂O (5g), Mg SO₄ x 7H₂O (0.25g), KCl (0.2g), (NH₄)₂SO₄ (0.1g), algal extract (25%) and Agar (15g). The pH was adjusted to 6.8 then the medium was autoclaved 120°C/20min.

2.3 Siderophores production

Siderophores production was carried out according to [19], but the ingredients are dissolved in Seawater instead of distilled water and added with algal extract of *U. lactuca* (25%).

3 Results

3.1 Growth on the medium with Seawater

Bacteria *Azospirillum brasilense* NH formed a subsurface growth pellicle 2-5mm deep in the semisolid medium containing algal extracts and the color was changed to blue brilliant, but there was non growth on the medium without algal extracts (control). On solid medium containing algal extract, small dense colonies appeared on the surface and non growth was shown on the solid medium without algal extracts (control). The growth in the both media semisolid and solid, confirm that the algal extract replaces all carbon source, such as malic acid.

3.2 Indole Acetic acid Production

A. brasilense NH produced high concentration of IAA (64.61 µg/ml) (Fig.1B) better than culture on NFB medium at 200 mM/l NaCl (55.78 g/ml) [5]. We should note that NH₄Cl and carbon source addition to the liquid medium for bacterial growth is not necessary (Figs.1 A).

3.3 Phosphate solubilization and siderophores production

Both of media showed positive results, NBRIP transparent hallos were appeared (figure 2A). The same result is obtained with siderophores, where yellow hallos around the spots are manifested (Figure 2B).

4 Discussion

A. brasilense NH growth in the medium containing only algal extracts and without malic acid or other carbon sources, lets us a conclude that *U. lactuca* provides the bacterial strain by carbon source in one hand, and by osmoprotectant to challenge the osmotic stress in the Seawater (32 g/l) in other hand. *A. brasilense* NH showed a best growth in presence of algal extracts at 600mM NaCl [5], and many bacteria like *E. coli* used the algal extracts as osmoprotectant [7]. It has been reported that marine macroalgae are rich in organic matter and nutrients.[15] showed that 1kg of dry mass of marine algae contained 7890 mg NH⁺ 1110-22000 mg total-N, 1370-13400 mg total-P. Bacteria can use the different

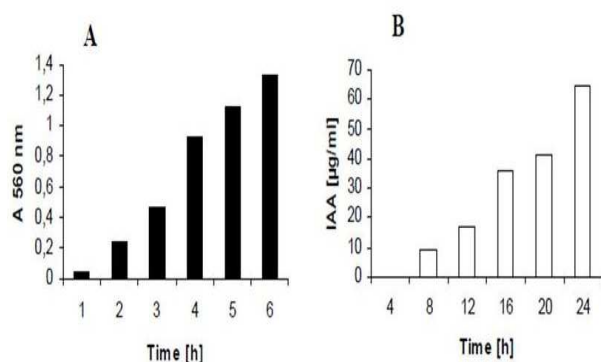


Fig. 1: Growth of *A. brasilense* NH liquid medium containing Seawater and evolution of IAA production.

A: growth (A560nm), B: IAA production (g/ml)

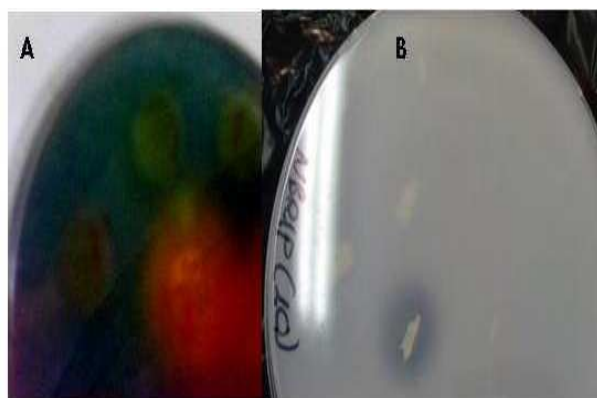


Fig. 2: Phosphatase and Siderophore production by *A. brasilense* NH

A: Siderophores and B: Phosphatase on NBRIP Medium.

nitrogen source and Cl^- presents in the water sea, so, the NH_4Cl should be added to the medium is replaced. Seawater is rich with mineral source replacing all mineral necessary for bacteria growing. Inoculation of the medium containing the water sea and algal extracts (10 and 20%) with *A. brasilense* NH does not show any growth (data not shown) it is due to the dominance of several minerals which can inhibit the growth of bacteria. Na^+ and Cl^- are very dominants in this environment, *A. brasilense* NH is known to use *Ulva lactuca* extract as osmoprotectant to challenge osmotic stress [13] and the marine alga has been reported to be rich in various betaines, amino-acids and dimethylsulphoniopropionate [7]. Different betaines are tested like osmoprotectants by *E. coli* [9,7], glycine betaine by *Azospirillum sp.* [8], and glycine betaine by *A. brasilense* NH [13]. The same case

for the dimethylsulphoniopropionate which has an osmoprotective effects on bacteria in Seawater (*Roseabacterdenitrificans*) [10].

Concerning production of high level of IAA in the same medium in presence of algal extracts in the absence of NH_4Cl like nitrogen source, and comparatively to NFB medium at 200mM NaCl [12], *A. brasilense* NH used algal extracts like osmoprotectants nitrogen source for the strain like as the growth. It has been reported [17] that IAA production it depends on the environmental conditions (temperature, carbon source and aeration). Algal extracts contain many carbon sources (amino-acids, betaines and DMSP).

5 Conclusion

These results demonstrated that *A. brasilense* NH get it natural environment in seawater in presence of algal extract. In the future it will be probable to use the Seawater to irrigation of wheat, especially in the arid soil messing water, because the strain has been shown that it promoted the wheat growth under saline condition in combination to *Ulva lactuca* in the previous study.

Acknowledgements

We express our deep acknowledgements for Prof. Dr. Anton Hartmann GSF-National Research Center for Environment and Health, Institute of Soil Ecology, Department of Rhizosphere Biology, Neuherberg / Munich, Germany, for his acceptance to teach many techniques in his laboratory, specially the study of IAA production, and to supervise my Phd thesis.

References

- [1] Aziz A, Martin-Tanguy J, Larher F, Plasticity of polyamine metabolism associated with high osmotic stress in rape leaf discs and with ethylene treatment. *Plant Growth Regul.* **21**, 153-163 (1997).
- [2] Barbier R, Galli E, Effect on wheat root 127 development of inoculation with an *Azospirillum brasilense* mutant with altered indole-3-acetic acid production, *Res. Microbiol.* **144**, 69-75 (1993).
- [3] Bashan Y and Holguin G, *Azospirillum*-plant relationships, environmental and physiological advances (1990-1996), *Can J. Microbiol.* **43**, 103-121 (1997).
- [4] Bashan Y, Holguin G, Bashan L, *Azospirillum* plant relationships: physiological, molecular, agricultural, and environmental advances (1997/2003). *Can J. Microbiol.* **50**, 521-577 (2004).
- [5] Bric JM, Bostock RM, Silverstone SE, Rapid in situ assay for indole acetic acid production by bacteria immobilization on a nitrocellulose membrane. *Appl. Environ. Microbiol.* **57**, 535-538 (1991).

- [6] Dobbelaere S, Vanderleyden J, Okon Y, Plant growth-promoting effects of diazotrophs in the rhizosphere. *CRC Critical Rev. Plant Sci.* **22**, 107-149 (2003).
- [7] Ghoul M, Minet J, Bernard T, Dupray E, Cornier M, Marine macroalgae as a source for osmoprotection for *Escherichia coli*. *Microb. Ecol.* **30**, 171-181 (1995).
- [8] Hartmann A, Prabhu SR, Galinski EA, Osmotolerance of diazotrophic rhizosphere bacteria. *Plant and Soil.* **137**, 105-109 (1991).
- [9] Lucht JM, Bremer E, Adaptation of *Escherichia coli* to high osmolarity environments: Osmoregulation of the high-affinity glycine betaine transport system ProU. *FEMS Microbiology reviews.* **14**, 3-20 (1994).
- [10] Ledyard KM, DeLong EF, Dacey JWH, Characterization of a DMSP-degrading bacterial isolate from the Sargasso Sea. *Archive of Microbiology.* 312-318 (1993).
- [11] Nabti E, Sahnoune M, Adjrad S, Van Dommelen A, Ghoul M, Schmid M, and Hofmann A, Rothballer M, Schmid M and Hartmann A. Enhancement and Restoration of growth of durum wheat (*Triticum durum* var. waha) on saline soil by using *Azospirillum brasilense* NH and marine alga *Ulva lactuca* In : *Algae : Ecology, Economic Uses and Environmental Impact.* Marine Biology (Edt) Nova Science Publishers, Inc, New York, 29-52 (2012).
- [12] Nabti E, Sahnoune M, Adjrad S, Van Dommelen A, Ghoul M, Schmid M, and Hofmann A, Rothballer M, Schmid M and Hartmann A. Restoration of growth of durum wheat (*Triticum durum* var. waha) under saline conditions due to inoculation with the rhizosphere bacterium (*Azospirillum brasilense* NH) and extracts of the marine alga (*Ulva lactuca*). *Journal of Plant Growth Regulation* **29**, 6-22 (2010).
- [13] Nabti E, Sahnoune M, Adjrad S, Van Dommelen A, Ghoul M, Schmid M, Hartmann A, A Halophilic and osmotolerant *Azospirillum brasilense* Strain from Algerian Soil restores Wheat Growth under saline Conditions. *Eng. Life Sci.* **4**, 354-360 (2007).
- [14] Nautiyal CS, An efficient microbiological growth medium for screening phosphate solubilizing microorganisms. *FEMS Microbiol. Lett.* **170**, 265-270(1999).
- [15] Nedzarek A, Rakusa-Suszczewski S, 2004. Decomposition of macroalgae and the release nutrient in Admiralty Bay, King Island, Antarctica. *Polar Biosc.* **17**, 26-35.
- [16] Okon Y, Vanderleyden J, Root-associated *Azospirillum* species can stimulate plantes. *ASM News.* **3**, 366-370 (1997).
- [17] Ona O, Van J, Prinsen E, Vanderleyden J, Growth and indole-3-acetic acid biosynthesis of *Azospirillum brasilense* Sp245 environmentally controlled, *FEMS Microbiology Letters.* **246**, 125-132 (2005).
- [18] Saxena B, Modi M, Modi V, Isolation and characterization of siderophores from *Azospirillum lipoferum* D-2. *J. Gen Microbiol.* **132**, 2219-2224 (1986).
- [19] Schwyn B, Neilands JB, Universal Chemical Assay for the Detection and Determination of siderophores. *FEMS Microbiol. Lett.* **10**, 110-119 (1986).
- [20] Seshadri S, Muthukumarasamy R, Lakshinarasimhan C, Ignacimuthu S, Solubilization of inorganic phosphates by *Azospirillum halopraeferans*. *Curr.Sci.* **79**, 565-567 (2000).