

Antagonistic Activity of Actinomyces Isolated from Azerbaijan's Soils

Behboodi Elham* and Hasanova Sevda

Department of Microbiology, Faculty of Biology, Baku State University, Azerbaijan.

Received: 15 Jun. 2016, Revised: 19 Jul. 2016, Accepted: 22 Jul.2016.

Published online: 1 Sep. 2016.

Abstract: Nowadays the idea of existence of acidophil actinomyces as a microbial community in soil does not raise doubt and they are one of the most important subjects of research. They have proved their own existence by growing in low-pH medium and formation of aerial mycelium and spores. In addition not only neutrophil but also acidophil actinomyces are considered to be the producers of antibacterial agencies and repressors of bacteria and fungi growth. They are famous for their second metabolic product (antibiotic). It has been proved that acidophil strains differ from neutrophil in their antagonistic property in relation to bacteria and fungi. This research compares the number and antagonistic property of neutrophil and acidophil actinomyces, isolated from Azerbaijan's soil.

Keywords: neutrophil, acidophil, actinomyces, antimicrobial activity.

1 Introduction

Among bacteria, the actinomycetes are important producers of bioactive compounds [1] and constitute a potential as biocontrol agents [2, 3, 4]. Actinomycetes represent a high proportion of soil microbial biomass, and appear to be of importance among the microbial flora of the rhizosphere [5]. Associations between actinomycetes and plant organs can be deleterious or beneficial for the host. While some actinomycetes secrete herbicidal compounds [6] or cause plant diseases [7], others can symbiotically fix atmospheric nitrogen [8] or protect plants against fungal infections (Cao et al. 2005). They protect to various degrees several different plants from soil-borne fungal pathogens [9, 10].

The possible existence of acidophilic and alkaliphilic, psychrophilic and thermophilic, halophilic and haloalkaliphilic, and xerophilic actinomycetes does not raise doubt in specialists [11, 12, 13, 14, 15]. The terms *acidophilic* and *acidotolerant* actinomycetes appeared in the literature at the turn of the last century [16,17]. The works presenting them have shaken the existed opinion that all actinomycetes are neutrophilic species [18]. Great number of acidophilic actinomycetes were representatives of the *Streptomyces* genus [19, 20, 21, 22]. Probably, this fact is related to their significant predominance in almost all the soils [11].

The major emphasis in the systematics of sporoactinomycetes has been on establishing relationships

between neutrophilic strains which, in culture, grow between pH 5.0 and 9.0 with an optimum close to neutrality [14, 23]. which share key chemotaxonomic and morphological properties with neutrophilic streptomycetes, grow in the range from about 3.5 to 6.5, with optimum rates at pH 4.5 to 5.5.

Streptomycetes have established themselves as the most potent group of microorganisms capable of forming a wide variety of antibiotics [24]. On the contrary, antifungal antibiotics effective against fungal disorders are relatively few mainly because of their solubility and toxicity problems [25, 26]. The ever increasing incidence of fungal infections in plants, animals and human being has directed the attention towards the search for actinomycetes producing novel antifungal antibiotics having a broad spectrum of activity and lesser toxicity [27, 28].

Indeed, different *Streptomyces* species produce about 75% of commercially and medically useful antibiotics [29]. According to [30], RhizovitR from *Streptomyces rimosus* is used in the control of a wide range of fungi such as *Pythium* spp., *Phytophthora* spp., *Rhizoctonia solani*, *Alternaria brassicicola*, and *Botrytis* sp. Liu et al. [31] also reported that *S. rimosus* showed a high antagonism activity against *Fusarium solani*, *F. oxysporium* f sp. *cucumarinum*, *Verticillium dahliae*, *R. solani*, *Fulvia fulva*, *Botrytis cinerea*, *A. alternat*, *Sclerotinia sclerotiorum* and

*Corresponding author e-mail: ebhboodi@gmail.com

Bipolarismaydis.

2 Materials and Methods

For screening and isolating of actinomycetes was used culture media Gause1 which is composed of: Starch (2%), K_2HPO_4 (0.05%), $MgSO_4$ (0.05%), NaCl (0.05%), KNO_3 (0.1%), $FeSO_4$ (0.001%), agar(2%). Actinomycetes colonies with different morphologies were selected and transferred to Gause1 slants for further studies.

2.1 Isolation actinomycetes from soil

Soil samples were collected from different regions of Azerbaijan: 4 Soil samples were analyzed in the first day - it is necessary to keep them in the cold place (in the fridge) during 2 days [13]. Samples (10g) of air-dried soil were mixed with sterile distilled water (100 ml) and then portion (1 ml) of soil suspensions (diluted 10^{-1}) were transferred to 9 ml of sterile distilled water and subsequently diluted to 10^{-2} , 10^{-3} , 10^{-4} , 10^{-5} and 10^{-6} and cultivated on the solid medium, were incubated at 30 °C for up to 20 days [32].

2.2 Characterization of the isolates

The selected *Actinomycetes* via antibacterial tests were characterized through morphological and biochemical tests. Morphological methods consisted of macroscopic and microscopic methods. The mycelium structure, color and arrangement of spores on the mycelium, and other properties such as the color of colonies, soil pH and etc. were observed. The observed structures were compared with Bergey's Manual of Determinative Bacteriology, Ninth edition (2000) and the organisms were identified. Moreover several biochemical tests such as Casein hydrolysis, starch hydrolysis and urea hydrolysis, acid production from various sugars, NaCl resistance and temperature tolerance were done.

2.3 Isolating in pure culture

To study physio-biochemical properties and bacterial development cycle and also determine their type, it is necessary to work with pure culture of microorganisms. For isolating to pure culture of aerobic microorganisms, it was seeded on Petri dishes from accumulated culture. Its drop was carried on the culture medium and rubbed by sterile spatula on the surface of medium, after this, by the same spatula were rubbed the other 3-4 dishes and incubated. Grown isolated colonies were taken out by loop to the test tube with solid sterile medium or in liquid medium [33].

3 Result and Discussion

3.1 Antimicrobial activity of isolated strains

About 160 strains of actinomycetes (129 neutrophil and 39 acidophil strains) were isolated from different types of

Azerbaijan's soils(Figure 1).All isolated acidophil and neutrophil actinomycetes belonged to Streptomyces genus. For subsequent analyze, 15 neutrophil and 15 acidophil actinomycetes were selected and their antimicrobial activity was studied by disk-diffusion method. The isolated cultures were screened against some microorganisms – *Staphylococcus aureus*, *Bacillus mezentericus*, *Escherichia coli*, *Aspergillusniger* and their antagonistic property was valued by appearance and measuring lysis zones around agar blocks of actinomycetes. The result is featured in table1 and figure 2.The analyses of antagonistic effect on test culture revealed considerable differences between acidophil and neutrophil soil actinomycestowards bacteria and fungi.

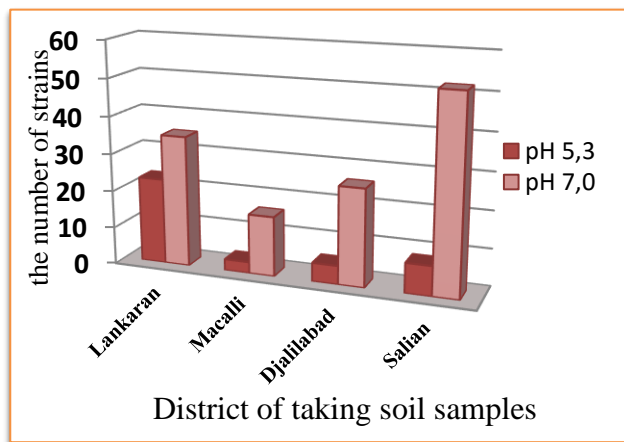


Fig.1: Distribution of actinomycetes in a neutral (pH 7, 0) and acidic medium (pH 5.3).

According to presented information on diagram (figure 1), the number of neutrophil strains in all soil samples were significantly more than acidophil strains. While the largest numbers of neutrophil actinomycetes were isolated from Saliian, the largest numbers of acidophil stains were isolated from Lankaran. In contrast the least numbers of acidophil and neutrophil actinomycetes were isolated from Masalli.

Table 1: The percentage of active acidophil and neutrophil actinomycete against test-cultures

Test –culture	pH 5.3	pH 7
<i>Staphylococcus aureus</i>	20%	26%
<i>Bacillus mezentericus</i>	40%	73%
<i>Escherichia coli</i>	53%	26%
<i>Aspergillusniger</i>	33%	13%

Among selected strains (neutrophil and acidophil) most of acidophil strains suppress the growth of *Escherichia coli* (53%) and they were less active against *Staphylococcus aureus* (20%). A great percentage of neutrophil strains (73%) show antagonistic property against *Bacillus mezentericus*. microscopic fungi *Aspergillusniger*, was more resistant against neutrophils, which just 13% of them prohibit its growth. While acidophil strains grown on the acidic medium proved to inhibit fungal growth [16] and gram negative *E.coli*, those grown on the neutral medium

suppress the growth of gram positive *St.aureus* and *B.mezentericus*.

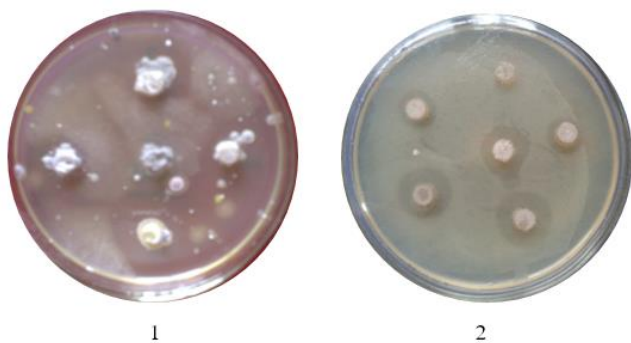


Fig.2: antagonistic property of acidophil (1) actinomycetes against *E.coli* and neutrophil (2) actinomycetes against *B.mezentericus*.

4 Conclusions

To conclude the number of neutrophil actinomycetes were considerably more than acidophils, although acidophils were more active against *Escherichia coli* and *Aspergillus niger*. It proves their antagonistic character in relation to fungi and bacteria.

References

[1] Lange L, Breinholt J, Rasmussen FW, Nielsen RI (1993). Microbial fungicides. the natural choice. *Pestic Sci.* 39:155–160.

[2] Kim BS, Moon SS, Hwang BK. (2000). Structure elucidation and fungal activity of an anthracycline antibiotic, daunomycin, isolated from *Actinomadura roseola*. *Agric Food Chem.* 48:1875–1881.

[3] Gomes RC, Seme`do LT, Soares RM, Alviano CS, Linhares LF, Coelho RR (2000). Chitinolytic activity of actinomycetes from acerrado soil and their potential in biocontrol. *LettApplMicrobiol* 30:146–150.

[4] Ouhdouch Y, Barakate M, Finace C (2001). Actinomycetes from Moroccan habitats. screening for antifungal activities. *Eur J Soil Biol.* 37:69–74.

[5] Sardi P, Saracchi M, Quaroni S, Petrolini B, Borgonovi GE, Nesli S (1992). Isolation of endophytic *Streptomyces* strains from surface-sterilized roots. *Appl Environ Microbiol.* 58:2691–2698.

[6] Tanaka Y, Omura S (1993). Agroactive compounds of microbial origin. *Annu Rev Microbiol.* 47:57–87.

[7] Locci R (1994). Actinomycetes as plant pathogens. *Eur J Plant Pathol.* 100:179–200.

[8] Oakley B, North M, Franklin JF, Hedlund BF, Staley JT (2004). Diversity and distribution of *Frankia* strains symbiotic with *Ceanothus* in California. *Appl Environ Microbiol.* 70:6444–6452.

[9] Xiao K, Kinkel LL, Samac DA (2002). Biological control of

Phytophthora root rots on alfalfa and soybean with *Streptomyces*. *Biol Control.* 23:285–295.

[10] El-Tarabily KA, Sivasithamparam K (2006). Non-streptomycete actinomycetes as biocontrol agents of soil-borne fungal plant pathogens and as plant growth promoters. *Soil Biol Biochem* 38:1505–1520.

[11] Zvyagintsev D. G. and Zenova G. M. (2001). *Ecology of Actinomycetes* (GEOS, Moscow).

[12] Zvyagintsev D. G. and Zenova G. M. (2002). *Actinomycetes in Saline and Alkaline Soils* (Knizhnyi Dom Universitet, Moscow). [in Russian].

[13] Zenova G. M. and Zvyagintsev D. G. (2002). *Diversity of Actinomycetes in Terrestrial Ecosystems* (Mosk.Gos.Univ., Moscow).

[14] Goodfellow M. and Williams S.T. (1983). Ecology of Actinomycetes. *Annu. Rev. Microbiol.* 37: 189–216.

[15] Jiang C. and L Xu. (1993). Actinomycetes Diversity in Unusual Habitats. *Actinomycetes* 4: 47–57.

[16] Khan M. R. and Williams S. T. (1975). Studies on the Ecology of Actinomycetes in Soil. Distribution and Characteristics of Acidophilic Actinomycetes. *Soil Biol. Biochem.* 7: 345–349.

[17] Williams S. T., Davies F. L., Mayfield C. I., and M. R. Khan. (1971). Studies on the Ecology of Actinomycetes in Soil: 11. The pH Requirements of Streptomyces from Two Acid Soils. *Soil Biol. Biochem.* 3:187–199.

[18] Kalakutskii L. V. and Agre N. S. (1977). *Development of Actinomycetes* (Nauka, Moscow) [in Russian].

[19] Goodfellow M. and Kim S. B. (1999). Phylogenetic Analysis of Acidophilic and Neutrotolerant Actinomycetes Isolated from Soil and Coal Wastes. in *The 11th International Symposium on the Biology of Actinomycetes*, Heraclion, Greece.

[20] Hagedorn C. (1976). Influence of Soil Acidity on Streptomyces Population in Habitat in Forest Soil. *Appl. Environ. Microbiol.* 32:358–375.

[21] Kim S. B., Seong C. N., Jeon S. J., et al. (2004). Taxonomic Study of Neutrotolerant Acidophilic Actinomycetes Isolated from Soil and Description of *Streptomyces yeochonensis* Sp. Nov. *Syst. Evol. Microbiol.* 54: 211–214.

[22] Park Y. H., Yim D. G., Kim E., et al. (1991). Classification of Acidophilic, Neutrotolerant, and Neutrophilic Streptomyces by Nucleotide Sequencing of 5s Ribosomal RNA. *Gen. Microbiol.* 137: 2265–2269.

[23] Kim B., Sahin N., Minnikin D.E., Zakrzewska-Czerwinska J., Mordarski M. and Goodfellow M. (1999). Classification of thermophilic streptomyces, including the description of *Streptomyces thermoalcalitolerans* sp. nov. *Syst. Bacteriol.* 7–17.

[24] Berdy, J. (1989). The discovery of new bioactive microbial metabolites: screening and identification. *Bioactive Metabolites from Microorganisms*. (Eds: Bushell, M. E., Grafe, U.) Elsevier, Amsterdam. 3-33.

[25] Bushell, M. E. (1982). Microbiological aspects of the discovery of novel secondary metabolites. *Topics in Enzyme and Fermentation Biotechnology* (Ed: Wiseman, A.). 6: 32-67.

- [26] Berdy, J. (1986).Further antibiotics with practical application.Biotechnology. (Eds: Pape, H., Rehm, H. J.) VCH, Weinheim, Deerfield Beach, FL. 4: 486-507.
- [27] Woodruff, H. B., Burg, R. W. (1986).The antibiotic explosion.Phannacological Methods, Receptors and Chemotherapy (Eds: Pamham, M. J., Bruinvels, J.) Elsevier, Amsterdam 303-351. Microbiol.
- [28] Georgiev, V. S. (1988).Fungal infections and the search for novel antifungal agents. Ann. N.Y Acad. Sci. 544: 1069-1076.
- [29] Miyadoh, S. (1993).Research on antibiotic screening in Japan over the last decade.a producing microorganisms approach. Actinomycetologica. 9:100–106.
- [30] Marten, P., Bruckner, S., Minkwitz, A., Luth, P., Bergm G. (2001).RhizovitR: Impact and formulation of a new bacterial product. In Koch, E., Leinonen P. (Eds.), Formulation of Microbial Inoculants: Proceedings of a meeting held in Braunschweig, Germany. COSTAction 830/Microbial inoculants for agriculture and environment, Germany, 78–82.
- [31] Liu, Q., Wu, Y.H., Yu, J.C.(2004a). Screening for antagonistic actionmyces isolates from greenhouse soil in northeast china. Soil. 36: 573–575 (in Chinese).
- [32] Звягинцев Д.Г. (1991).Методы почвенной микробиологии и биохимии. МГУ, москва.
- [33] Зенова Г.М., Степанов А.Л., Лихачева А.А., Манучарова Н.А., (2002).Практикум по биологии почв. МГУ, москва.
- [34] Теппер Е.З., Шильникова В.К, Переверзева Г.И. (1979). Практикум по биологии. Второе издание. Москва.
-