

Effect of Acetylation on Physico-chemical Characteristics of *Sweitenia mycrophylla* Gum: A Potential Excipient.

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Abstract: Recent studies focus on the chemical modification of polysaccharides for biomedical applications. Chemical modification of these polysaccharides produces products with improved physicochemical and functional properties that are not available from commercial polysaccharides. The modification discussed includes acetylation of *Sweitenia mycrophylla* gum with acetic anhydride in the presence of sodium hydroxide. The resulting product was characterized by FTIR and NMR spectroscopy. The degree of acetylation is 0.24. Physicochemical characteristics such as solubility, viscosity and swelling index of the native *Sweitenia mycrophylla* gum and acetylated gum were also determined. The results showed that the acetylated gum had higher values of solubility, viscosity and swelling index as 93.40% at 80°C, 65.4cs and 46.54% respectively while the native gum had solubility viscosity and swelling index as 30.10% at 80°C, 28.40cs and 15.20% respectively. Chemical modification via acetylation increased the solubility, viscosity and swelling index of *Sweitenia mycrophylla* gum. The experimental work provides enough evidence to exploit this natural biopolymer in food, textile and pharmaceutical industry, especially as an efficient alternative approach for the oral delivery of hydrophilic macromolecules.

Keywords: *Sweitenia mycrophylla* Gum, chemical modification, acetyl group, drug delivery.

1 Introduction

In recent years, the development and utilization of polysaccharides isolated from natural sources have attracted increasing attention in biochemistry, pharmacology and food chemistry, due to their sustainability, biodegradability and biosafety [1]. *Sweitenia mycrophylla* gum polysaccharide is an exudate extracted from the bark of monogamy tree. *Sweitenia mycrophylla* tree is a large tree, reaching a height of 30-40m. Gum is produced from cuts at the bark of the tree for sales in markets in Bombay, India [34]. Due to excellent properties of gums such as solubility, viscosity, thickening, binding, stabilizing and emulsifying, they are utilized in several multibillion-dollar industries such as adhesive, cosmetic, confectionaries, paint, paper, pharmaceutical and most importantly Food [3, 24-25]. Even if gum and its derivatives are well known for a wide range of applications, like other polysaccharides, there are evidence of some drawbacks, such as uncontrolled rate of hydration, pH-dependency, solubility, thermal decomposition, low shear stress resistance, high retrogradation and syneresis. [1, 4]. Chemical modification

provides an efficient route not only for removing such drawbacks but also for improving physicochemical properties such as solubility, viscosity and swelling index and to introduce new properties for different applications. A number of modifications via chemical treatment can be effected resulting in products suitable for specific purposes in the food and pharmaceutical industries [5]. According to [2] chemical modification of *anacardium occidentale* gum by oxidation increases the uronic acid content of the gum from 3.7% to 38% which further increases solubility and water holding capacity. Also according to [6], oxidation of gum generally increases their hydrophilicity and solution clarity which make them more soluble in aqueous system. Chemical modification through acetylation generally increases the emulsifying capacity which further increases swelling index and solubility [7]. Nowadays, the development of new products in gum based industries are searching for gums with different or better physicochemical and functional properties such as viscosity, solubility, low retrogradation and syneresis tendency. In recent years, substantial progress have been made in obtaining polysaccharides from non-conventional botanical sources

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and studying their functional and physicochemical properties [8-11].

The acetylated gum is produced by the esterification of native gum with acetyl groups. The efficiency of the reaction is affected by factors such as reagent concentration reaction time, pH, and presence of catalyst and gum source. The functional properties of the gum acetate will depend on the number of acetyl group incorporated to the sugar unit of gum molecules [12 -14].

Acetylation was selected as a chemical means of attaching pendant acetic anhydride groups due to its technical simplicity, low cost of chemical reagents and wide range applications to produce acetylated gum. The result of this research is likely to highlight the effect of acetylation on the physicochemical properties of *Sweitenia mycrophylla* gum in order to amplify the possibilities of the gum applications in food and pharmaceuticals as an emulsifier, effective binder and suspending agent in drug formulation.

The objective of this research is to prepare and characterize acetylated *Sweitenia mycrophylla* gum in order to improve on its physicochemical characteristics and amplify the possibilities of the gum applications.

2 Materials and Methods

Sweitenia mycrophylla gum was collected by tapping in March, 2010 from owena forestry Ondo –State, Nigeria was identified and authenticated at the herbarium of the department of plant science technology, university of Jos, Nigeria. The plant superficial incision was made at the bark of the tree and the bark was later stripped off. After five weeks, gum was manually collected. The gum samples were dried at room temperature, cleaned, milled with Kenwood blender [UK], sieved through a mesh-size 250 microns to obtain fine – size particles, kept in labeled plastic container and stored in the refrigerator for subsequent analysis.

3 Purification of Gum Sample

Dried crude gum [10g] was stirred in cold distilled water [250ml] for 3 hours at room temperature. The supernatant was obtained by centrifugation. The supernatant was made up to 500ml and treated with ethanol [1.4v/v] in order to precipitate the carbohydrate. The material was washed again with ethanol followed by distilled water and freeze-dried.

3.1 Preparation of acetylated gum

Acetylated gum was obtained using the method reported by [13]. In brief, gum (10g) was dispersed in 50cm³ of distilled water and then constantly stirred for 30 minutes. The slurry was adjusted to pH 8.0 with 3% NaOH. Acetic anhydride (1.2g) was then added to the slurry. After the addition of

the acetic anhydride, the reaction was allowed to proceed for another five minutes. The pH of the slurry was adjusted to 4.5 with 0.5M HCl and filtered through whatman No 1 filter paper. The residue obtained was washed four times with distilled water to remove completely some acids that may be present in the product and finally air dried at room temperature.

3.2 Solubility

The solubility of gum was determined according to a standard method reported by [15]. Gum sample (10g) was suspended in 40ml of distilled water. It was heated to the desired temperature (60⁰c, 70⁰c and 80⁰c) for 30 minutes with continuous shaking. The mixtures were centrifuged at 1,000rpm for 15 minutes. An aliquot of supernatant (5ml) was evaporated at 130⁰c and weighed. The percentage solubility of the gum was the ratio in mass (g) of the dried supernatant to the initial mass (g) of the dry gum expressed in percent.

3.3 Swelling and gel fraction studies

Swelling and gel fraction studies were carried out according to a standard method reported by [16] Samples weighing 0.01g of gum were placed in small dishes that were carefully inserted into glass flasks. Total volume of 60mL distilled water was slowly poured into each glass flasks. The samples were allowed to soak for 2 hours at room temperature, after which the excess solution was carefully removed and the galled sample remaining in the gelled bottle were weighed. The galled samples were lyophilized for three days and then weighed again. The swelling ratio and percentage of gel fraction were calculated. Using Equations (1) and (2)

$$\text{Swelling ratio} = W_{\text{water}}/W_{\text{gel}} \quad (1)$$

$$\text{Percentage fraction} = W_{\text{gel}}/W_{\text{solid}} \times 100 \quad (2)$$

W_{water} = weight of the sample after 2 hours soaking

W_{gel} = weight of the sample after lyophilization

W_{solid} = initial weight of the sample.

3.4 Viscosity

Apparent viscosity of gum was determined using a Brookfield Viscometer (Model RVF, Stoughton, MA). The gum slurry (5%) was placed in a boiling bath for 15 minutes and then cooled to 22⁰c. cold paste viscosity was determined using spindle at 25⁰c.

3.5 Fourier Transform Infra-Red (FTIR) Spectroscopy

FTIR spectra were obtained on a FTIR spectrometer [Shumadzu 8400s] using a KBr disc. The equipment was operated with a resolution of 4cm⁻¹ and the scanning range

from 4000 to 400cm⁻¹

3.6 Nuclear magnetic resonance (NMR) spectroscopy

¹³C NMR spectra were recorded in an NMR (600MHz) spectrometer (Agilent Technology, America). The sample (10mg) was dissolved in 700μL at 70⁰ C with continuous stirring for 6 hours followed by sonication for 10 minutes. The solution was centrifuged and transferred to a 5mm NMR tube. Chemical shift were reported in ppm relative to internal standard TMSP.

3.7 Statistical analysis

The data obtained from the study were analyzed using the Statistical Analysis System (SAS) software and the means were separated by T-Test.

4 Result and Discussion

Table1: Physicochemical characteristics of acetylated and native *Sweitenia mycrophylla* gum.

		Solubility (%)	Swelling ratio (%)	Viscosity (cs)	Gel fraction (%)
Native gum	60 ⁰ c	3.90 ± 0.01			
	70 ⁰ c	23.10 ± 0.04			
	80 ⁰ c	30.10 ± 0.03	15.2 ± 0.4	28.40 ± 0.30	60 ± 4.8
Act gum	60 ⁰ c	46.70 ± 6.05			
	70 ⁰ c	80.80 ± 5.14	46.1 ± 0.2	65.20 ± 1.4	34 ± 2.81
	80 ⁰ c	93.40 ± 7.20			

Mean ± S.D, n = 3. Act=acetylated

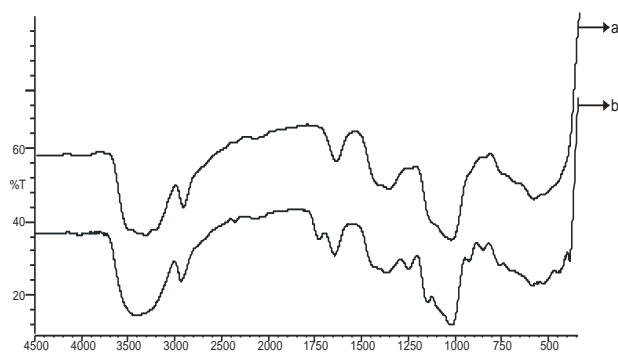


Fig 1: FTIR of (a) unmodified *Sweitenia mycrophylla* gum (b) Acetylated *Sweitenia mycrophylla* gum.

The viscosity value for the acetylated gum (65.2 ± 1.40 cs) (Table1) was higher than the native gum (28.40 ± 0.30 cs). This higher value of viscosity could be explained by the increase in the swelling power and solubility of the chemically modified gum. According to [17], acetylation of gum generally increases the emulsifying capacity which further increases the viscosity and water holding capacity. During the acetylation process, the gum-gum interactions in the granules are weakened by the introduction of acetyl groups, this makes the gum to be more attracted towards water molecules [18-20, 26].

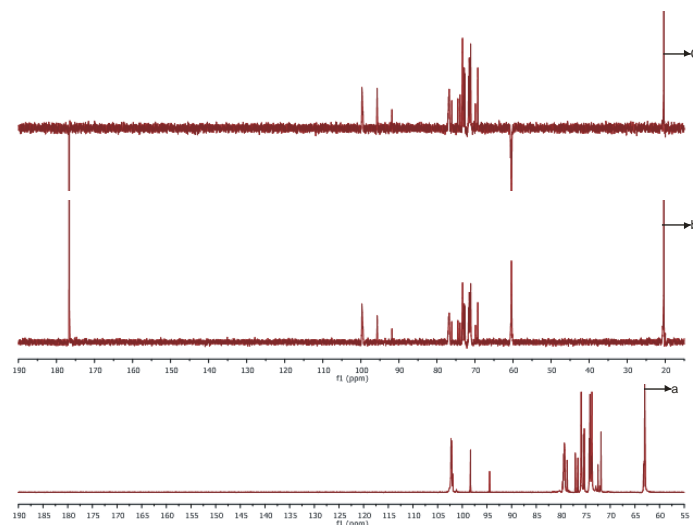


Fig. 2: ¹³C NMR Spectra obtained for (a) Unmodified *Sweitenia mycrophylla* gum (b) acetylated *Sweitenia mycrophylla* gum (c). ¹³C-DEPT for acetylated *Sweitenia mycrophylla* gum.

Also the swelling index and solubility of acetylated gum were higher than the native gum (Table1). Hovers and susilki.[21] reported that the introduction of acetyl groups during the acetylation process reduces the bond strength between gum molecules (amylose/ amylopectin) and thereby increases the swelling power and solubility of the gum granules. This facilitates access of water to amorphous areas, enhancing the water holding capacity of the gum matrix and developing a more organized structure, leading to a higher resistance to deformation and achieving a higher peak viscosity.

The solubility of modified and unmodified gum profoundly increased with increase temperature (Table 1). At temperature above 90⁰c, more than 100% modified gum were dissolved. This is due to introduction of acetyl groups which is bulkier than hydroxyl groups and capable of obstructing chain association. The superior solubility and swelling index of acetylated gum compared with the native gum may be due to the presence of hydrophilic substituting groups (CH₃C=O) which allow the retention of water molecules because of their ability to form hydrogen bonds. [22-23]. However, the swelling ratio of the acetylated carbohydrate increases while the gelling properties reduces, which is the main reason for studying the gel fraction percentage. The FTIR spectra of the native and acetylated gum is shown in Fig.1 The polysaccharide unit of sugar with hydroxyl group [OH] as the major functional groups appear in the region [3650 to 3200cm⁻¹] and disappeared when the gum was acetylated, there was introduction of acetic groups and the spectra now processed peaks around [1750cm⁻¹ to 1735cm⁻¹] attributed to C=O stretching, indicating the presence of the acetyl group. This peak was seen to decrease and in the case of native gum the peak was not resolved from that of the C=O group. The peak at 3300-3400cm⁻¹ caused by OH stretching was also seen to

decrease with an increase in acetyl content.

In the ^{13}C NMR spectroscopy, the carbon anomeric region shows two major signals which assigned as C-1 of α -D-galactose (residue A) at 98.87ppm and C-1 of β -D-mannose (residue B) at 102.1ppm. The resonances of the carbon atoms were well resolved (Fig 2a) and identified as the resonances of C-2, C-3, C-4 and C-5 (77.2, 73.5, 77.1, 75.2ppm) respectively for residue B and C-2, c-3, C-4, and C-5 (71.90, 73.0, 74.80, 76.00 ppm) for residue. A. This facts are almost identical with gums of other origin [31, 32, 33].

Spectrum for the acetylated gum (Fig. 2b) shows some differences in relations to unmodified gum. The anomeric signals decrease considerably due to sugar residues, probably because of chain degradation [31]. A new signal at 179.4ppm and 20.37ppm were observed for acetylated gum in comparison with the unmodified gum. In ^{13}C DEPT NMR spectrum (fig 2c), the signal at 62.5ppm (CH_2) appeared with the opposite amplitude to those of CH_3 and CH , which can be attributed to the acetylation of CH_2 primary carbons (C-6). The presence of acetylated group causes an increase in the ^{13}C chemical shift.

The absence of signal inversion at 20.37ppm of the acetylated gum spectrum (fig 2c) demonstrated the correct signal assignment of the CH_3 of acetyl group introduced while the inverted signal at 179.4ppm shows the presence of $-\text{COO}$ groups with no hydrogen atoms [31, 32, 33]. Furthermore, the shift of the peaks of the C-6 of A and B carbon atoms to 61.18ppm from 60.44 and 60.92ppm indicated the position on the carbohydrate ring where substitution occurred, however, the peaks were not sufficiently resolved to show separate peaks for the acetylation at position of C-6 of residue A and B. Thus, the FTIR and NMR spectra confirms the acetylation of *Sweitenia mycrophylla* gum.

5 Conclusion

The study confirms that purification by extraction and acetylation may improve physicochemical properties of *sweitenia mycrophylla* gum the feasibility of the procedure was demonstrated by FTIR and NMR spectroscopy. Furthermore, the obtained product can have wider biological application as drug delivery carriers by grafting/crosslinking compounds of interest.

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